

Short communication

Synthesis of new benzimidazole–coumarin conjugates as anti-hepatitis C virus agents

Jih Ru Hwu^{a,*}, Raghunath Singha^a, Shih Ching Hong^a, Yung Hsiung Chang^a,
Asish R. Das^a, Inge Vliegen^b, Erik De Clercq^b, Johan Neyts^b

^a Department of Chemistry, National Tsing Hua University, Hsinchu 30013, Taiwan, ROC

^b Rega Institute for Medical Research, Katholieke Universiteit Leuven, Minderbroedersstraat 10, B-3000 Leuven, Belgium

Received 8 March 2007; accepted 12 September 2007

Abstract

Nineteen new conjugated compounds were successfully synthesized by a one-flask method from benzimidazole and coumarin derivatives. A methylenethio linker was used to connect these two kinds of derivatives. In addition, substituted benzimidazol-2-thiones were also coupled with β -D-glucose peracetate; the resultant glucosides were further converted to the corresponding 2-(methylthio)coumarin derivatives. Their activity against the hepatitis C virus was tested; two of the most potent compounds 2-[(6'-bromocoumarin-3'-yl)methylenethio]-5-fluorobenzimidazole (**4i**) and its derivative 1-[(2'',3'',4'',6''-tetra-*O*-acetyl)glucopyranos-1''-yl]-2-[(6'-bromocoumarin-3'-yl)methylenethio]benzimidazole (**7c**) showed EC₅₀ values of 3.4 μ M and 4.1 μ M, respectively. At a concentration of 5.0 μ M, compound **7c** inhibited HCV RNA replication by 90% and had no effect on cell proliferation. Given these data, a structure–activity relationship was established.

© 2007 Elsevier B.V. All rights reserved.

Keywords: Benzimidazole; Coumarin; Conjugate; *N*-Glucosides; Anti-HCV

About 3% of the current world population (~170 million people) are chronically infected by the chronic hepatitis C virus (HCV), which causes a progressive liver disease (Lauer and Walker, 2001). The infection can lead to lethal complications, such as cirrhosis and hepatocellular carcinoma (Penin et al., 2004). Liver failure from chronic hepatitis C is the leading cause for liver transplantation in the West (Armstrong et al., 2000; Condrón et al., 2005). Current therapies result in only 50–60% of the patients in a sustained virological response (Bretner, 2005a). There is thus an unmet need for potent antiviral compounds (Pawlotsky and Gish, 2006; Neyts, 2006).

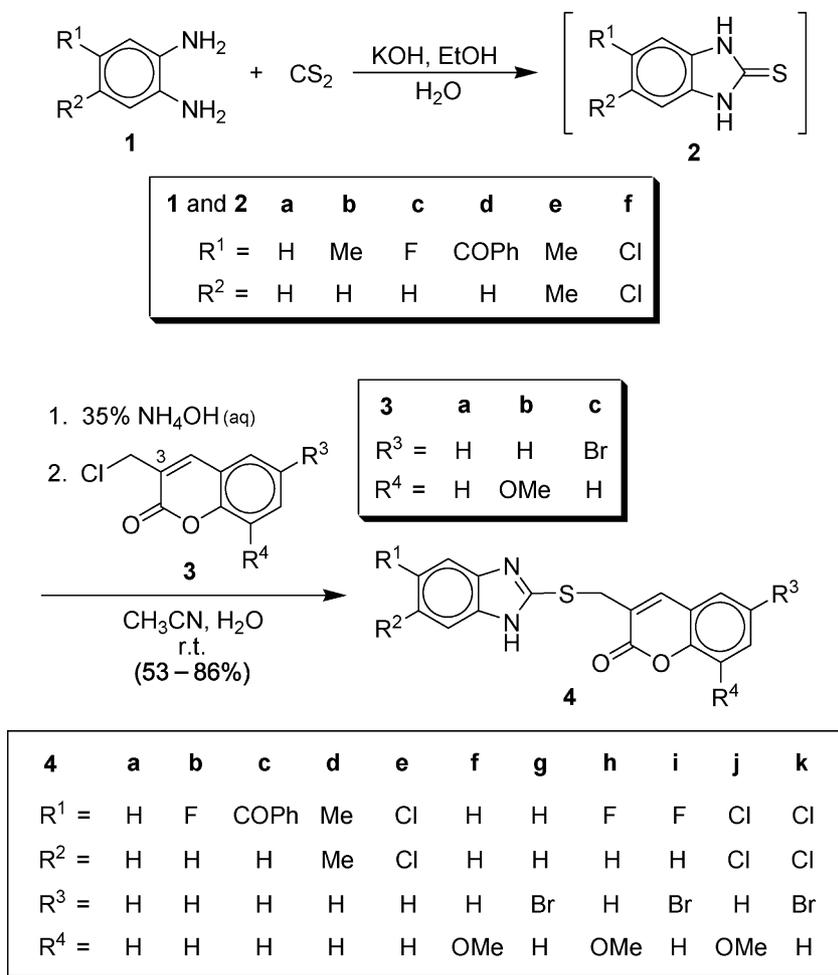
Some benzimidazole (Andrzejewska et al., 2002; Gümüs et al., 2003) and coumarin (Ito et al., 2003; Nam et al., 2002) derivatives show diverse biological activities with significant clinical potential, including the treatment of leukaemia (Demirayak et al., 2002; Garuti et al., 2000) and cancer (Antonini et al., 1988; Lukevics et al., 2001). These compounds may also possess potent antiviral activity (Curini et al., 2003; Devivar et al., 1994). Examples include a series of non-nucleoside benzimidazoles

reported by Beaulieu et al. (Beaulieu et al., 2004a,b; Beaulieu and Llinàs-Brunet, 2002; McKercher et al., 2004). Recently 2-arylbenzimidazole-5-carboxylic acids were shown to inhibit the HCV NS5B RNA polymerase (Hirashima et al., 2006; Ishida et al., 2006). To the best of our knowledge, hybridized molecules containing benzimidazole and coumarin have never been reported as anti-HCV agents (cf. Bretner et al., 2005b). We here report that benzimidazole–coumarin conjugates with a methylenethio linker and the corresponding *N*-glucosides exhibit antiviral activity against HCV.

1. Synthesis of new benzimidazoles 4a–k and the corresponding glucopyranosides 7a–e

We treated various substituted phenylenediamines **1a–f** with carbon disulfide (Klimesová et al., 2002) and ethanolic KOH in H₂O to give intermediates **2a–f** (see Scheme 1). Subsequently, aqueous NH₄OH (35%) and 3-chloromethylcoumarins **3a–c** were added in situ to afford the desired benzimidazole–SCH₂–coumarin derivatives **4a–k**, respectively, which were purified by HPLC. Their overall yields were 53–86% and purity was >99.8% as checked by GC and HPLC.

* Corresponding author. Tel.: +886 35 725813; fax: +886 35 721594.
E-mail address: jrhwu@mx.nthu.edu.tw (J.R. Hwu).

Scheme 1. A “one-flask” method for the synthesis of benzimidazole–SCH₂–coumarin conjugates.

Structures of the conjugated products were identified by spectroscopic methods. For example, the ¹H NMR spectrum of conjugate **4i** showed a singlet at 4.33 ppm for the SCH₂ protons and a singlet at 7.95 ppm for the CH=C–COO proton in the coumarin moiety. Its ¹³C NMR spectrum exhibited resonance at 30.66 ppm for the SCH₂ carbon and 149.94 ppm for the –N=C(–N)(–S) carbon. For the elemental analysis of **4i**, the calculated percentages are C 50.50, H 2.49, Br 19.54, F 4.70, N 6.93, O 7.92, S 7.91; which were found C 50.39, H 2.53, Br 19.48, F 4.66, N 6.97, O 7.89, S 7.86.

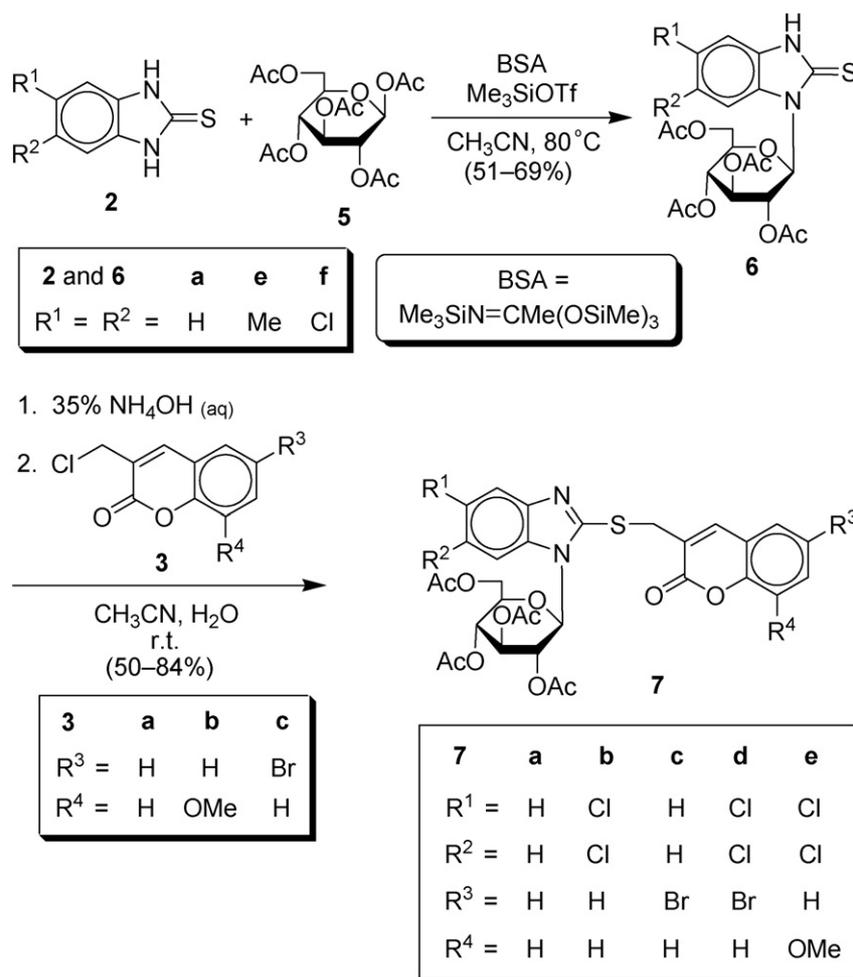
Our design by using the sulfur atom in the –SCH₂– linker to attach the benzimidazole ring directly was to facilitate tautomerization of the imidazole nucleus. Consequently, the regioisomeric problem for R¹ ≠ R² in compound **4** can be circumvented during the preparation of **4b**, **4c**, **4h**, and **4i**.

Xu et al. (Duan et al., 2004) isolated *O*-galloyl-β-D-glucoses from traditional Chinese medicine *Galla Chinese*. Three glucose esters in this family are identified as inhibitors of hepatitis C virus NS3 protease. Consequently we considered to synthesize glucosides **7** by coupling the heterocycles **4** with a glucose ester. Unfortunately, direct coupling of conjugated heterocycles **4a–k** with *O*-peracetylpyranose **5** met with failure. Therefore, we decided to silylate 2-thiones **2** with *N,O*-

bis(trimethylsilyl)acetamide (BSA) (Bhan et al., 1997; Khodair et al., 2003); the intermediates were then coupled with *O*-peracetylpyranose **5** in the presence of Me₃SiOTf at 80 °C (Scheme 2).

In the reactions **2** + **5** → **6**, a mixture of regioisomers might be produced when unsymmetrical 2-thiones **2b–d** were used as the starting materials. To circumvent the potential problems of separation and structural identification, we adopted the symmetrical benzimidazole-2-thiones **2a**, **2e**, and **2f** to react with pyranose **5** individually. Accordingly, compounds **6a**, **6e**, and **6f** were isolated as the exclusive products. These glucosidic 2-thiones were alkylated with various 3-(chloromethyl)coumarins **3a–c** in the presence of aqueous NH₄OH (35%) (Khodair et al., 2003) at room temperature to give the desired *N*-glucopyranosyl conjugates **7a–e** with purity >99.7%, which were purified and checked by HPLC.

The structures of **7a–e** were satisfactorily characterized by spectroscopic methods. For example, the high-resolution mass of glucoside **7d** was found 783.9896, which is consistent with the theoretical value 783.9899 for C₃₁H₂₇BrCl₂N₂O₁₁S. Its ¹³C NMR spectrum showed 31 peaks for every carbon atoms therein in full. In the ¹H NMR spectrum, the glycosidic proton resonated at the 5.55 ppm as a doublet with *J* = 8.4 Hz, which indicates

Scheme 2. A synthetic pathway to generate *N*-glucosides of benzimidazole–SCH₂–coumarin conjugates.

the β configuration of glycoside (Zhang et al., 2005). The two ClCH₂–hydrogens in the starting material **3c** at 4.53 ppm as a singlet was shifted to 4.40 ppm and 4.84 ppm as two doublets with $J = 11$ Hz for the two diastereotopic SCH₂ hydrogens in the product **7d**. Appearance of these peaks with an AB pattern indicates the success of the coupling reaction **6f** + **3c** \rightarrow **7d**. Furthermore, the elemental analysis results of **7d** are listed below: the calculated percentages are C 47.45, H 3.47, Br 10.07, Cl 8.92, N 3.57, O 22.44, S 4.08; they were found C 47.51, H 3.44, Br 9.99, Cl 8.88, N 3.61, and S 4.11.

2. Antiviral activity tests

We performed antiviral assays with the 19 synthesized conjugated compounds against 15 viruses in four different cell lines. They included HIV-1_{IIIIB} and HIV-2_{ROD} in MT-4 cells; herpes simplex virus-1 (KOS), thymidine kinase-deficient (TK⁻), herpes simplex virus-2 (G), vaccinia virus, and vesicular stomatitis virus in E₆SM cells; respiratory syncytial virus in HeLa cells; as well as parainfluenza-3 virus, reovirus-1, Sindbis virus, Coxsackie B4 virus, and Punta Toro virus in Vero cell cultures. None of the compounds proved active against any of the aforementioned viruses.

3. Antiviral evaluation in the hcv genotype 1b subgenomic replicon

The potential antiviral activity of the compounds was initially evaluated in the Huh 5-2 replicon system, which was kindly provided by Prof. R. Bartenschlager, University of Heidelberg, Germany (Lohmann et al., 1999). The antiviral assays and cytostatic determination assays have been described in detail before (Paeshuyse et al., 2006). Among the 19 newly synthesized benzimidazole–coumarin conjugates, some were found to inhibit HCV subgenomic replicon replication in the Huh 5-2 cell line. The 50% inhibitory concentrations for viral replication (EC₅₀) and host cell growth (CC₅₀), as well as the selectivity indices (SI = CC₅₀/EC₅₀), are presented in Table 1. Conjugate **4i** and glucoside **7c** inhibited HCV replication with EC₅₀ values of 3.4 μ M and 4.1 μ M, respectively. To exclude that the observed antiviral effect was caused by inhibition of the luciferase activity in the replicon assay, or to particular characteristics of this Huh 5-2 replicon construct, we further explored the antiviral activity of compounds **4i** and **7c** by means of a quantitative RT-PCR assay in three other HCV replicon models (Coelmont et al., 2006; Paeshuyse et al., 2006). The activity of the compounds was confirmed in these assays (see Table 2).

Table 1
Inhibitory effects of conjugated compounds **4**, **6**, and **7** on HCV subgenomic replicon replication in Huh 5-2 cells

Compound ^a	R ¹	R ²	R ³	R ⁴	CC ₅₀ (μM) ^{b,c}	EC ₅₀ (μM) ^{c,d}	SI ^e
4a	H	H	H	H	90	27	3.4
4b	F	H	H	H	45	11	3.9
4c	COPh	H	H	H	37	6.7	5.6
4d	Me	Me	H	H	67	15	4.4
4e	Cl	Cl	H	H	74	15	4.8
4f	H	H	H	OMe	27	10	2.8
4g	H	H	Br	H	42	4.0	10
4h	F	H	H	OMe	42	16	2.6
4i	F	H	Br	H	27	3.4	8.0
4j	Cl	Cl	H	OMe	11	3.6	3.1
4k	Cl	Cl	Br	H	44	2.3	19
6a	H	H	H	H	104	104	1.0
6e	Me	Me	H	H	98	100	0.98
6f	Cl	Cl	H	H	60	26	2.3
7a	H	H	H	H	76	11	6.7
7b	Cl	Cl	H	H	20	3.1	6.3
7c	H	H	Br	H	43	4.1	10
7d	Cl	Cl	Br	H	>64.8	4.1	>16
7e	Cl	Cl	H	OMe	33	4.3	7.6

^a Interferon α-2b was used as a (positive) reference compound at 10,000 units/well and reduced the signal in the viral RNA (luciferase) assay to background levels without any cytotoxic activity.

^b Cytostatic concentration: the concentration required to reduced cell proliferation by 50%.

^c The values obtained as the average of triplicate determinations.

^d Effective concentration: the concentration required to inhibit HCV RNA replication by 50%.

^e Selectivity index (ratio of CC₅₀ to EC₅₀).

A dose–response effect of **7c** on HCV RNA replication in Huh 5-2 cells and of their effect on host cell proliferation is depicted in Fig. 1. For example, at a concentration of 5.0 μM, compound **7c** inhibited HCV RNA replication by 90%, whereas at this concentration it had almost no effect on cell proliferation. At a concentration of 16 μM, compound **7c** inhibited HCV replicon replication by 99% whereas cell growth was reduced only by ~20%. These data clearly indicate that the antiviral activity is specific because potent inhibition was observed at concentrations that have no, or little, effect on the cell proliferation.

4. Structure–activity relationship

Different substituents including F, Cl, Br, Me, OMe, and COPh were placed at various positions of the benzimidazole–

SCH₂–coumarin conjugates (see **4a–k**). By analyzing the EC₅₀ and CC₅₀ values shown in Table 1, we can deduce the following structure–activity relationships: (a) introduction of an F-, Cl-, or Me-substituent on the benzimidazole ring (i.e., **4b**, **4e** and **4d** versus **4a**) showed little effect on their HCV inhibitory activity; (b) introduction of a Br group on the coumarin ring (e.g., **4g** versus **4a**) enhanced HCV inhibition by 6.7-fold and also the selectivity by 2.9-fold; (c) introduction of an OMe group on the coumarin ring (e.g., **4j** versus **4e**) further improved the antiviral activity; (d) enhancement of the selectivity resulting from substituents in the coumarin nucleus (cf. **4f**, **4a**, and **4g**) followed the order OMe < H < Br; (e) attachment of the –CH₂–coumarin moiety onto the benzimidazole-2-thione nucleus connected with a pyranose (e.g., **6a** versus **7a**) enhanced

Table 2
Inhibitory effects of conjugated compounds **4i** and **7c** on HCV subgenomic replicon replication in Huh 6, Huh 9-13, and Huh mono cells

Cell	Compound	CC ₅₀ (μM) ^a	EC ₅₀ (μM) ^b	SI ^c
Huh 6	4i	23 ± 6.0	3.0 ± 1.6	7.7
Huh 6	7c	25 ± 7.2	6.0 ± 0.8	4.2
Huh 9-13	4i	26 ± 3.6	4.6 ± 1.0	5.7
Huh 9-13	7c	73 ± 2.9	0.8 ± 0.5	91
Huh mono	4i	26 ± 6.0	9.0 ± 8.0	2.9
Huh mono	7c	31 ± 14	12 ± 7.0	2.6

^a Cytostatic concentration: the concentration required to reduced cell proliferation by 50%. Data are mean values from two independent experiments.

^b Effective concentration: the concentration required to inhibit HCV RNA replication by 50%. Data are mean values from two experiments.

^c Selectivity index (ratio of CC₅₀ to EC₅₀).

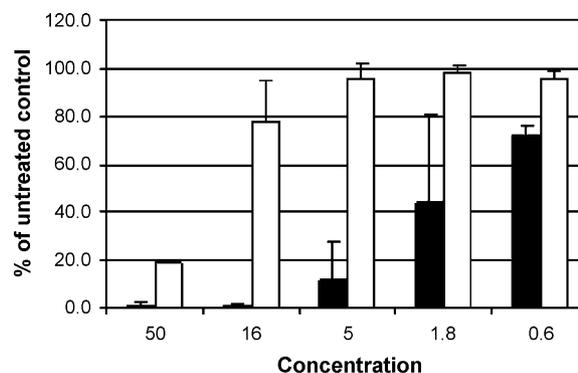


Fig. 1. Dose–response effect of compound **7c** on HCV subgenomic replication and cell proliferation in Huh 5-2 cells. Black bars represent the effect on HCV replicon content, open bars the effect on host cell proliferation. Data are mean values ± S.D. for two independent experiments.

the HCV inhibitory activity by >9.4-fold and also the selectivity by 6.7-fold; (f) incorporation of a β -D-glucose peracetate moiety into the benzimidazole–coumarin conjugate (e.g., **4e** versus **7b**) resulted in a 4.8-fold increase in anti-HCV activity.

In conclusion, a series of substituted benzimidazole–coumarin conjugated compounds were synthesized and evaluated for their activities against various viruses. In this compound library, several new conjugates resulted in an inhibitory effect on HCV replication: the conjugate **4i** displayed an EC₅₀ of 3.4 μ M. The *N*-glucoside **7c** inhibited HCV RNA replication by 90% and 99% at concentrations of 5.0 μ M and 16 μ M, respectively. These conjugates may be considered as potential lead anti-HCV compounds for further selectivity optimization.

Acknowledgements

For financial support, we thank the National Science Council (Grant No. 92-2751-B-007-002-Y) and Ministry of Education of R.O.C. The work in Leuven was supported by the Fonds voor Wetenschappelijk Onderzoek Vlaanderen (G.0267.03) and by the VIRGIL European Network of Excellence on Antiviral Drug Resistance grant (LSHM-CT-2004-503359) from the Priority 1 “Life Sciences, Genomics and Biotechnology for Health” program in the 6th framework program of the EU. We thank Mrs. Katrien Geerts for fine experimental assistance.

References

- Andrzejewska, M., Yépez-Mulia, L., Cedillo-Rivera, R., Tapia, A., Vilpo, L., Vilpo, J., Kazimierczuk, Z., 2002. Synthesis, antiproteoal and anticancer activity of substituted 2-trifluoromethyl- and 2-pentafluoroethylbenzimidazoles. *Eur. J. Med. Chem.* 37, 973–978.
- Antonini, I., Claudi, F., Cristalli, G., Franchetti, P., Grifantini, M., Martelli, S., 1988. Heterocyclic quinones with potential antitumor activity. 2. Synthesis and antitumor activity of some benzimidazole-4,7-dione derivatives. *J. Med. Chem.* 31, 260–264.
- Armstrong, G.L., Alter, M.J., McQuillan, G.M., Margolis, H.S., 2000. The past incidence of hepatitis C virus infection: implications for the future burden of chronic liver disease in the United States. *Hepatology* 31, 777–782.
- Beaulieu, P.L., Llinàs-Brunet, M., 2002. Therapies for hepatitis C infection: targeting the non-structural proteins of HCV. *Curr. Med. Chem. Anti-Infect. Agents* 1, 163–176.
- Beaulieu, P.L., Bös, M., Bousquet, Y., DeRoy, P., Fazal, G., Gauthier, J., Gillard, J., Goulet, S., McKercher, G., Poupart, M.-A., Valois, S., Kukulj, G., 2004a. Non-nucleoside inhibitors of the hepatitis C virus NS5B polymerase: discovery of benzimidazole 5-carboxylic amide derivatives with low-nanomolar potency. *Bioorg. Med. Chem. Lett.* 14, 967–971.
- Beaulieu, P.L., Bousquet, Y., Gauthier, J., Gillard, J., Marquis, M., McKercher, G., Pellerein, C., Valois, S., Kukulj, G., 2004b. Non-nucleoside benzimidazole-based allosteric inhibitors of the hepatitis C virus NS5B polymerase: Inhibition of subgenomic hepatitis C virus RNA replicons in huh-7 cells. *J. Med. Chem.* 47, 6884–6892.
- Bhan, P., Bhan, A., Hong, M., Hartwell, J.G., Saunders, J.M., Hoke, G.D., 1997. 2',5'-Linked oligo-3'-deoxyribonucleoside phosphorothioate chimeras: thermal stability and antisense inhibition of gene expression. *Nucl. Acids Res.* 25, 3310–3317.
- Bretner, M., 2005a. Existing and future therapeutic options for hepatitis C virus infection. *Acta Biochim. Pol.* 52, 57–70.
- Bretner, M., Baier, A., Kopanska, K., Najda, A., Schoof, A., Reinholz, M., Lipniacki, A., Piasek, A., Kulikowski, T., Borowski, P., 2005b. Synthesis and biological activity of 1*H*-benzotriazole and 1*H*-benzimidazole analogues—inhibitors of the NTPase/helicase of HCV and of some related Flaviviridae. *Antivir. Chem. Chemother.* 16, 315–326.
- Coelmont, L., Paeshuyse, J., Windisch, P., De Clercq, M., Bartenschlager, E., Neyts, R.J., 2006. Ribavirin antagonizes the in vitro anti-hepatitis C virus activity of 2'-*C*-methylcytidine, the active component of valopicitabine. *Antimicrob. Agents Chemother.* 50, 3444–3446.
- Condron, S.L., Heneghan, M.A., Patel, K., Dev, A., McHutchison, J.G., Muir, A.J., 2005. Effect of donor age on survival of liver transplant recipients with hepatitis C virus infection. *Transplantation* 80, 145–148.
- Curini, M., Epifano, F., Maltese, F., Marcotullio, M.C., Gonzales, S.P., Rodriguez, J.C., 2003. Synthesis of collinin, an antiviral coumarin. *Aust. J. Chem.* 56, 59–60.
- Demirayak, S., Mohsen, U.A., Karaburun, A.C., 2002. Synthesis and anticancer and anti-HIV testing of some pyrazino[1-*a*]benzimidazole derivatives. *Eur. J. Med. Chem.* 37, 255–260.
- Devivar, R.V., Kawashima, E., Revankar, G.R., Breitenbach, J.M., Kreske, E.D., Drach, J.C., Townsend, L.B., 1994. Benzimidazole ribonucleosides: design, synthesis, and antiviral activity of certain 2-(alkylthio)- and 2-(benzylthio)-5,6-dichloro-1-(β -D-ribofuranosyl)benzimidazoles. *J. Med. Chem.* 37, 2942–2949.
- Duan, D., Li, Z., Luo, H., Zhang, W., Chen, L., Xu, X., 2004. Antiviral compounds from traditional Chinese medicines *Galla Chinese* as inhibitors of HCV NS3 protease. *Bioorg. Med. Chem. Lett.* 14, 6041–6044.
- Garuti, L., Roberti, M., Malagoli, M., Rossi, T., Castelli, M., 2000. Synthesis and antiproliferative activity of some benzimidazole-4,7-dione derivatives. *Bioorg. Med. Chem. Lett.* 10, 2193–2195.
- Gümüş, F., Algül, Ö., Eren, G., Eroglu, H., Diril, N., Gür, S., Özkul, A., 2003. Synthesis, in vitro cytotoxic and antiviral activity of *cis*-[Pt(R(-) and S(+)-2- α -hydroxybenzylbenzimidazole)₂Cl₂] complexes. *Eur. J. Med. Chem.* 38, 473–480.
- Hirashima, S., Suzuki, T., Ishida, T., Noji, S., Yata, S., Ando, I., Komatsu, M., Ikeda, S., Hashimoto, H., 2006. Benzimidazole derivatives bearing substituted biphenyls as hepatitis C virus NS5B RNA-dependent RNA polymerase inhibitors: structure–activity relationship studies and identification of a potent and highly selective inhibitor JTK-109. *J. Med. Chem.* 49, 4721–4736.
- Ishida, T., Suzuki, T., Hirashima, S., Mizutani, K., Yoshida, A., Ando, I., Ikeda, S., Adachi, T., Hashimoto, H., 2006. Benzimidazole inhibitors of hepatitis C virus NS5B polymerase: identification of 2-[(4-diarylmethoxy)phenyl]-benzimidazole. *Bioorg. Med. Chem. Lett.* 16, 1859–1863.
- Ito, C., Itoigawa, M., Mishina, Y., Filho, V.C., Enjo, F., Tokuda, H., Nishino, H., Furukawa, H., 2003. Chemical constituents of *Calophyllum brasiliense* 2. Structure of three new coumarins and cancer chemopreventive activity of 4-substituted coumarins. *J. Nat. Prod.* 66, 368–371.
- Khodair, A.I., Al-Masoudi, N.A., Gesson, J.-P., 2003. A new approach to the synthesis of benzothiazole, benzoxazole, and pyridine nucleosides as potential. *Antitumor Agents Nucleosides Nucl. Acids* 22, 2061–2076.
- Klimesová, V., Kocí, J., Pour, M., Stachel, J., Waisser, K., Kaustová, J., 2002. Synthesis and preliminary evaluation of benzimidazole derivatives as antimicrobial agents. *Eur. J. Med. Chem.* 37, 409–418.
- Lauer, G.M., Walker, B.D., 2001. Hepatitis C virus infection. *N. Engl. J. Med.* 345, 41–52.
- Lohmann, V., Korner, F., Koch, J., Herian, U., Theilmann, L., 1999. Barten-schlager R. Replication of subgenomic hepatitis C virus RNAs in a hepatoma cell line. *Science* 285, 110–113.
- Lukevics, E., Arsenyan, P., Shestakova, I., Domracheva, I., Nesterova, A., Pudova, O., 2001. Synthesis and antitumor activity of trimethylsilylpropyl substituted benzimidazoles. *Eur. J. Med. Chem.* 36, 507–515.
- McKercher, G., Beaulieu, P.L., Lamarre, D., LaPlante, S., Lefebvre, S., Pellerin, C., Thauvette, L., Kukulj, G., 2004. Specific inhibitors of HCV polymerase identified using an NS5B with lower affinity for template/primer substrate. *Nucl. Acids Res.* 32, 422–431.
- Nam, N.-H., Kim, Y., You, Y.-J., Hong, D.-H., Kim, H.-M., Ahn, B.-Z., 2002. Preliminary structure–antiangiogenic activity relationships of 4-seneciolyloxymethyl-6,7-dimethoxycoumarin. *Bioorg. Med. Chem. Lett.* 12, 2345–2348.
- Neyts, J., 2006. Selective inhibitors of hepatitis C virus replication. *Antivir. Res.* 71, 363–371.

- Paeshuyse, J., Kaul, A., De Clercq, E., Rosenwirth, B., Dumont, J.M., Scalfaro, P., Bartenschlager, R., Neyts, J., 2006. The non-immunosuppressive cyclosporin DEBIO-025 is a potent inhibitor of hepatitis C virus replication in vitro. *Hepatology* 43, 761–770.
- Pawlotsky, J.M., Gish, R.G., 2006. Future therapies for hepatitis C. *Antivir. Ther.* 11, 397–408.
- Penin, F., Dubuisson, J., Rey, F.A., Moradpour, D., Pawlotsky, J.-M., 2004. Structural biology of hepatitis C virus. *Hepatology* 39, 5–19.
- Zhang, G., Shen, J., Cheng, H., Zhu, L., Fang, L., Luo, S., Muller, M.T., Lee, G.E., Wei, L., Du, Y., Sun, D., Wang, P.G., 2005. Syntheses and biological activities of rebeccamycin analogues with uncommon sugars. *J. Med. Chem.* 48, 2600–2611.